

Subscriber access provided by ISTANBUL TEKNIK UNIV

A Cytotoxic #-Carboline from the Bryozoan Catenicella cribraria

John A. Beutler, John H. Cardellina II., Tanya Prather, Robert H. Shoemaker, Michael R. Boyd, and Kenneth M. Snader

J. Nat. Prod., 1993, 56 (10), 1825-1826• DOI: 10.1021/np50100a026 • Publication Date (Web): 01 July 2004

Downloaded from http://pubs.acs.org on April 4, 2009

More About This Article

The permalink http://dx.doi.org/10.1021/np50100a026 provides access to:

- Links to articles and content related to this article
- Copyright permission to reproduce figures and/or text from this article



Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

A CYTOTOXIC β-CARBOLINE FROM THE BRYOZOAN CATENICELLA CRIBRARIA

John A. Beutler, John H. Cardellina II, Tanya Prather, Robert H. Shoemaker, Michael R. Boyd,*

Laboratory of Drug Discovery Research & Development

and KENNETH M. SNADER

Natural Products Branch, Developmental Therapeutics Program, Division of Cancer Treatment, National Cancer Institute, Building 1052, Frederick, Maryland 21702-1201

ABSTRACT.---1-Vinyl-8-hydroxy- β -carboline was identified as the cytotoxic constituent of the bryozoans *Catenicella cribraria* and *Cribricellina cribraria*. Literature nmr data for this previously known compound, now reported from a new source, were corrected.

Compared to other major groups of marine invertebrates, the bryozoans remain a relatively poorly examined phylum in the realm of marine natural products chemistry (1). Extracts of two bryozoans, Catenicella cribraria Busk (phylum Bryozoa, class Gymnolaemata, order Cheilostomata, family Vittaticellidae) and Cribricellina cribraria Busk (phylum Bryozoa, class Gymnolaemata, order Cheilostomata, family Mucronellidae), showed relatively potent, but modestly differential, cytotoxicity in the NCI 60cell human tumor assay. Bioassay-guided fractionation provided a single cytotoxic compound, 1-vinyl-8-hydroxy-βcarboline, previously isolated from the latter species (2). No previous work on the chemistry of Cat. cribraria has been reported.

Solvent-solvent partitioning of the crude organic extracts of each bryozoan collection yielded cytotoxic CHCl₃ fractions. Normal phase diol and amino bonded phase hplc provided a pure compound, which we characterized as 1-vinyl-8-hydroxy- β -carboline. Comparison of nmr spectral data with the literature values (2) for 1-vinyl-8-hydroxy- β -carboline disclosed several discrepancies which could not be explained solely by the marked pH dependence of chemical shifts in MeOH- d_4 . Erroneous transcription of data into the table of Prinsep *et al.* (2) explained the remaining discrepan-

cies (J.W. Blunt, personal communication).

The previously reported cytotoxicity of 1-vinyl-8-hydroxy- β -carboline was paralleled by the relatively potent cytotoxicity observed in the NCI human tumor 60-cell-line assay. The very modest differential cytotoxicity produced by the crude extract carried through to the purified compound; the most sensitive cell line subpanel was the melanoma subpanel, where the LC_{50} values of eight of nine cell lines were similar to or slightly less than the mean panel LC_{50} . The mean panel response parameter values for 1-vinyl-8hydroxy- β -carboline were 1.8 μ M at the GI₅₀, 5.8 μ M at the TGI level, and 19 μ M at the LC₅₀ level.

EXPERIMENTAL

COLLECTIONS.—*Cat. cribraria*, an orange, erect, branching bushy colony, was collected at a depth of 14 m on March 15, 1989, on the underside of a rock substrate overhang, 0.6 km southwest of the light on Cape Vlamingh, Rottnest Island, Western Australia.

Cr. cribraria, an orange branching colony, was collected at a depth of 15 m on February 17, 1987, on the inside vertical rock walls of a tunnel at Poor Knights Island, The Tunnel, North Wall, New Zealand.

Voucher specimens are on deposit at the Smithsonian Institution.

EXTRACTION.—Frozen whole bryozoans were ground with dry ice in a hamburger mill, and the dry ice was allowed to sublime. The thawed tissue was stirred with distilled H_2O at 3° for 3 h and filtered to generate an aqueous extract, and the marc was lyophilized. The dry marc was extracted with CH_2Cl_2 -MeOH (1:1) overnight and rinsed with MeOH, and the solvent was removed with a rotary evaporator at 35° to yield an organic extract. *Cat. cribraria* (136 g wet wt) yielded 1.95 g of organic extract (1.4%). *Cr. cribraria* (238 g wet wt) yielded 4.29 g of organic extract (1.8%).

The crude extracts of Cat. cribraria (224 mg) and Cr. cribraria (232 mg) were subjected to a solvent-solvent partitioning scheme (3). In each case, cytotoxicity was concentrated in the CHCl₃ fraction; tlc indicated the same major Dragendorffpositive spot in each fraction. The CHCl₃ fractions of each extract were separately filtered through 3 g of diol chromatography media in MeOH and then subjected to hplc on diol media (YMC 2×25 cm, 60 Å, 5 μ), using a gradient of EtOAc to MeOH and monitoring by diode array uv detection. The major uv-absorbing peak in both CHCl₃ fractions was collected and found to have identical 'H- and ¹³C-nmr spectra. HMBC, HMQC, difference nOe, uv, and hr fabms characterization of this compound gave a structure identical to that of 1-vinyl-8-hydroxy-β-carboline; however, the H-nmr spectral data were not identical to those reported in the literature (2). Communication with the senior authors of the prior work (2) revealed an error in the transcription of ¹H-nmr data. Our data were in close agreement with their actual ¹H-nmr data: δ 8.22(d, J=5.4, H-3), 7.93(d, 5.3, H-4), 7.62(dd, 5.3, H-4), 7.62(dd, 5.3, H-4))8, 1, H-5), 7.45 (dd, 17.4, 11, H-1'), 7.07 (dd, 8, 7.6, H-6), 6.96 (dd, 7.6, 1, H-7), 6.39 (dd, 17.4, 1.4, H₂-2'), 5.65 (dd, 11, 1.4, H_b-2').

BIOASSAYS.—The 60-cell-line human disease-oriented tumor screening panel, its operation, and data presentation have been described previously (4). Tracking assays for the isolation procedure employed the previously described XTT tetrazolium methodology (5) and the LOX IMVI melanoma and U251 CNS tumor cell lines selected from the NCI panel.

ACKNOWLEDGMENTS

We thank A. Monks and D. Scudiero for the 60-cell line panel assays, T. McCloud for the extractions, M. Munro of the University of Canterbury, New Zealand and P. Murphy of the Australian Institute of Marine Sciences for the collections, and G. Gray for mass spectra. We appreciate J. Blunt's assistance in resolving the discrepancies in the nmr data of 1.

LITERATURE CITED

- U. Anthoni, P.H. Nielsen, M. Pereira, and C. Cristophersen, *Comp. Biochem. Physiol. B*, 96B, 431 (1990).
- M.R. Prinsep, J.W. Blunt, and M.H.G. Munro, J. Nat. Prod., 54, 1068 (1991).
- S.H. Grode, T.R. James Jr., J.H. Cardellina II, and K.D. Onan, J. Org. Chem., 48, 5203 (1983).
- M.R. Boyd, in: "Cancer: Principles and Practice of Oncology Updates." Ed. by V.T. DeVita Jr., S. Hellman, and S.A. Rosenberg, Lippincott, Philadelphia, 1989, Vol. 3, no. 10, pp. 1–12.
- D.A. Scudiero, R.H. Shoemaker, K.D. Paull, A. Monks, S. Tierney, T.H. Nofziger, M.J. Currens, D. Seniff, and M.R. Boyd, *Cancer Res.*, 48, 4827 (1988).

Received 4 February 1993